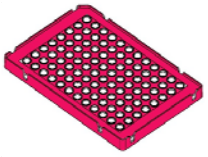
# ProfilerPro Glycan Profiling Quick Guide

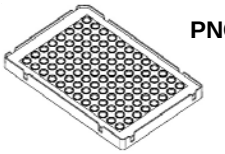
## Sample Preparation

**Denature**
1. Thaw and spin Denaturing Plate at 1200g for 1 minute.
2. Carefully remove plate seal.
3. Add 8µL of sample (monoclonal antibody) with concentration range of 1.25mg/mL to 7.5mg/mL (10µg to 60µg total protein) to Denaturing Plate. Mix by pipetting up and down or with a plate shaker.
4. Seal plate carefully with an adhesive plate seal.
5. Spin plate at 1200g for 1 minute.
6. Incubate for 10 minutes at 70C using a PCR machine or heat block.



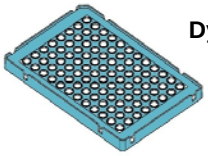
**Denaturing Plate**

**Digestion**
1. Thaw PNGase F Plate.
2. Spin both PNGase F Plate and Denaturing Plate at 1200g for 1 minute.
3. Carefully remove plate seals.
4. Transfer all denatured sample to PNGase F Plate. Mix by pipetting up and down or with a plate shaker.
5. Seal plate carefully with an adhesive plate seal.
6. Spin PNGase F Plate at 1200g for 1 minute.
7. Incubate for 1 hour at 37C using a PCR machine or heat block.



**PNGase F Plate**

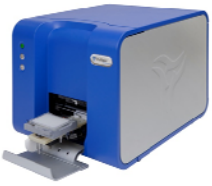
**Labeling**
1. Thaw Dye Plate.
2. Spin both Dye Plate and PNGase F Plate at 1200g for 1 minute.
3. Carefully remove plate seals.
4. Transfer 8µL of Digested sample to the Dye Plate. Mix by pipetting up and down or with a plate shaker.
5. Spin Dye Plate at 1200g for 1 minute.
6. Incubate the unsealed plate for 2 hours at 55C, or until dry using a PCR machine (lid open) or heat block.



**Dye Plate**

**Reconstitution**
1. Add 100µL of molecular grade water to dried samples.
2. Seal plate carefully with an adhesive plate seal.
3. Mix samples on a plate shaker at maximum speed for at least 1 minute.
4. Spin plate at 1200g for 1 minute.
5. Carefully remove plate seal.
6. Run plate on LabChip GXII.

**LabChip GXII**



For full Profiler Pro Glycan Profiling User Guide, go to
www.caliperls.com/chip-datasheets

Caliper LifeSciences




PN: 450708 Rev3

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## Chip Preparation Procedures

*Note: Allow the chip and Gel Matrix to equilibrate to room temperature for about 20-30 minutes before use.*

### Preparing the Buffer Tube

1. Add 750µL of molecular grade water to the 0.75mL Buffer Tube.
2. Insert the Buffer Tube into the buffer slot on the LabChip GXII instrument.

Buffer Tube

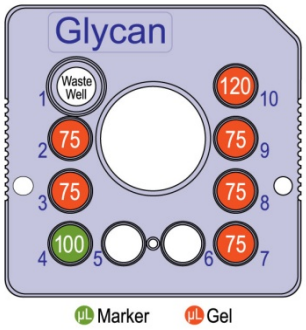
### Preparing the Ladder Tube

1. Add 145µL of Ladder Diluent to one of the Ladder tubes.
2. Vortex at highest speed for about 30 seconds and spin down.
3. Transfer 120µL of prepared ladder to the 0.2mL Ladder Tube.
4. Insert the Ladder Tube into the ladder slot on the LabChip GXII instrument.

Ladder Tube

### Preparing the Chip

1. Remove reagents from all wells of the chip using a vacuum.
2. Rinse and aspirate all wells with molecular grade water. Repeat, and be careful to aspirate all the water in the wells and any water that may have spilled onto the outside of the chip.
3. Add 75µL of Gel Matrix to chip wells 2, 3, 7, 8 and 9 and 120µL in well 10 using a Reverse Pipetting Technique.
4. Prepare marker solution by adding 125µL of Marker Diluent to one of the Marker tubes. Vortex at highest speed for about 30 seconds and spin down. Transfer 100µL of prepared marker solution to chip well 4. (*Note: Prepare marker solution just before loading the chip in the LabChip GXII and starting the assay. Do not prepare marker solution in advance as the marker signal degrades over time.*)
5. Place the chip in the LabChip GXII instrument to begin the assay.

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